

Performance Comparison of Image Normalisation Method for DNA Microarray Data

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ABSTRACT

Normalisation is a process of removing systematic variation that affects measured gene expression levels in microarray experiment. The purpose is to get a more accurate DNA microarray result by deleting the systematic errors that may have occurred when making the DNA microarray slide. In this paper, four normalisation methods of Global, Lowess, Quantile and Print-tip are discussed, tested and their final results compared in the form of Matrixes and graphs. Ideal and real microarray slides have been used for this project. It was found that the Print-tip normalisation method showed the closest results to the real result for an ideal microarray slide and it has a straight median line final graph. The Print-tip normalisation method uses more than one normalization factor that is divided among intervals which are dependent on the values of the addition of red and green logarithm.

Keywords: DNA, Microarray, Normalisation, Global, Lowess, Quantile, Print-tip, Background correction, M-A plot

INTRODUCTION

Gene expression measurements provide clues on the regulatory mechanism, biochemical pathways and broader cellular function. By

gene expression is the transformation process of gene's information into proteins. The formal transformational pathway of protein begins with the DNA (deoxyribonucleic acid) which is copied to the mRNA (messenger ribonucleic acid) and, finally this molecule passes from nucleus to cytoplasm carrying the information to build proteins (Belean et al., 2011).

There are many microarray analysis software packages in the market. Each software program is concerned with three main tasks: 1) gridding or addressing, which is

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the process of specifying coordinate to every spot on the slide 2) segmentation which decides the classification of each pixel either as foreground which corresponds to be an interest spot or as background which acts as an error or noise 3) Intensity Extraction which is the step to calculate green and red for foreground fluorescence intensity for each spot on the array (Borda et al., 2011; Rao et al., 2008).

Processes to inspect the results and correct the errors are: 1) background correction method obtained by subtracting the value of the background intensity from the value of foreground intensity or any other suitable method to neglect the effect of background intensity 2) normalisation method which is the objective of this research (Yang et al., 2001).

Normalisation is the process of removing systematic variations that affect measured gene expression levels in microarray experiments. The purpose of normalisation is to adjust for effects which arise from variations in the microarray technology rather than from biological differences between the RNA samples or between the printed probes. Imbalances between the red and green dyes may arise from differences between the labelling efficiencies or scanning properties of the two dyes complications perhaps by the use of different scanner settings (Geeleher et al., 2009). The aim of this paper is to review various methods that discuss and compare DNA microarray normalization.

In section II several normalization algorithms are elaborated, while section IV discusses the comparison of these various methods. Section V and VI presents the methodology and results of analysis of the different methods. The conclusion follows in section VII.

LITERATURE REVIEW

Discussion on the normalisation of DNA microarray is currently well developed. Before we review some of them, we will explain the two types of graphs that can show normalisation quality. First, (log M vs. log R) as shown in Figure 1(a). Second, M-A plot is 45° rotation of standard scatter plot as shown in Figure 1(b). Write R and G for the background-corrected red and green intensities for each spot. Normalisation is usually applied to the log-ratios of expression, which will be written ($M = \log R - \log G$). The log-intensity of each spot will be written ($A = (\log R + \log G)/2$), a measure of the overall brightness of the spot. (The letter M is a mnemonic for minus while A is a mnemonic for addition) (Dudoit et al., 2002).

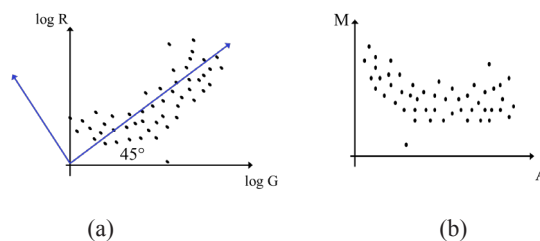


Figure 1. (a) Log R vs. Log G; (b) M-A Plot

This section will discuss and elaborate the methods of DNA microarray normalisation and identify the most suitable for further microarray analysis. The first method is Global normalisation: the underlying assumption of this approach is that the total of mRNA labelled with either R value (sum of red intensities) or G value (sum of green intensities) is equal. While the intensity for any one spot may be higher in one channel than the other, when averaged over thousands of spots in the array, these fluctuations should average out. In this method, the value of c out of $\log(R/G)$. The c value is equal to the main assumption that equal to \log of the total R over total G which can be expressed by the variable K (Yang et al., 2002). The intensity-dependent lowess normalisation runs a line through the middle of the MA plot, shifting the M value of the pair (A, M) by $c=c(A)$, as shown in Equation 3. One estimate of $c(A)$ is made using the loess function: Locally Weighted Scatterplot Smoothing (Berger et al., 2004; Bilban et al., 2002).

$$\log_2 R/G \rightarrow \log_2 R/G - c = \log_2 R/(kG) \tag{1}$$

$$K = \sum R/G \tag{2}$$

$$\log_2 R/G \rightarrow \log_2 R/G - c(A) = \log_2 R/(k(A)G) \tag{3}$$

In the Print-tip normalisation, each M -value ($\log R - \log G$) is normalised by subtracting from it the corresponding value of the tip group loess curve that is dependent on A value ($[\log R + \log G]/2$) while its value should be fixed. The normalised \log -ratios (N) are the residuals from the tip group loess regressions. A simpler form of Print-tip is shown in Equation 4 where $\text{loess}(A)$ is the global loess curve plotted in Figure 2. Refer to Figure 3 for the final figure of the Print-tip normalisation (Smyth et al., 2003). The Quantile normalisation method is undertaken by rearranging the genes in each column as in second table in Figure 4. Following which the mean in each row is replaced the whole row by the mean value as shown in the third table in Figure 4. Finally, reorder each gene in its original place with its new value.

$$N = M - \text{loess}(A) \tag{4}$$

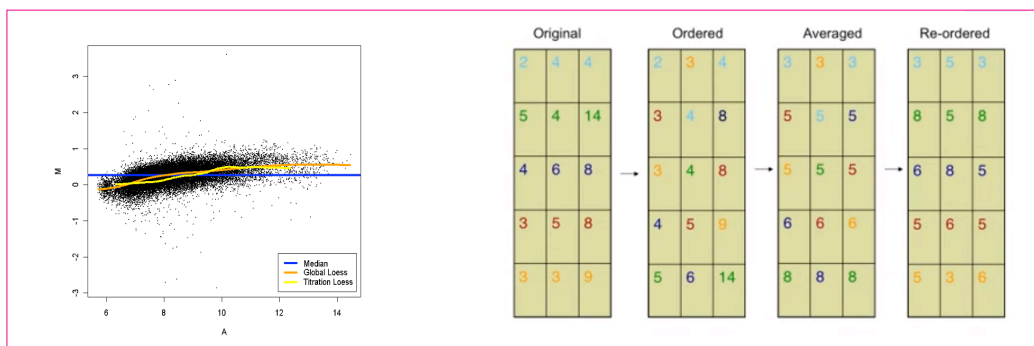


Figure 2. Global normalisation

Figure 3. Print-tip normalisation; Figure 4. Quantile normalisation

COMPARISON OF DIFFERENT NORMALISATION APPROACHES

In this section, the existing system algorithm as discussed in section III will be analysed and discussed to find out the similarities and variations among the different normalisation methods. Table 1 summarized the comparison of these algorithms.

From Table 1, it can be seen that, all the methods are using mainly the value of M which equal to log of red intensity minus log of green intensity. However, three methods have different value to subtract from M. To illustrate, Global normalisation use the log of addition of each of red and green intensity while the other two methods are using median and global median.

In term of the final shape of the normalisation on M-A graph, there are similarities between Lowess and Print-tip methods because both have a straight median line in the value of (M=0) due to their similarities on subtracting the mean or median from M. However, in Global normalisation, there is a curve around the value of (M=0) due to the subtraction of the total R and G. Quantile normalisation method does not use M-A plot, consequently its final graphs do not always take a straight line of the mean on the (M=0). According to this review, we suggest Print-tip normalisation method to be used because when comparing to the global normalisation its final figure is simpler and easier to read and can also easily be compared to various plots. Straight line on (M=0) is easier to read than the Global and lowess normalisation curve.

Table 1
Comparison between different system algorithms

No.	[1]	[2]	[3]	[4]
Method	Global	Lowess	Print Tip	Quantile
Function	Log (R/ KG)	Log (R/ G) – c(A)	N= M- loess (A)	Mean of rows after reorder
Variable	$K = \sum R/G$	LOWESS function	Global Loess	NA
Shape on M-A graph	Curve	Straight line on (M=0)	Straight line on (M=0) but has some variation	It does not meet M-A plot.

METHODOLOGY

Using Matlab, we developed a code that can extract the intensity for 100 spots. Using 100 spots instead of the whole microarray slide make the process easier and simpler especially to compare the many algorithms used. In order to examine the suitable method which would be more accurate and suitable for this project, an ideal microarray image spots in Figure 5(a), and a real microarray slide in Figure 5(b) were used. Matlab usually reads the image intensity as matrix by pixel, for example our image after cropping is 220*227 pixels while it has only 100 spots. Thus, each spot has around 20 pixel diameters. Next, it calculates the foreground and background then subtract the background value from foreground, and using threshold equal to zero will not allow negative values to appear. In the ideal image the value of background is fixed (Rb = Gb =3) while foreground value is a variant from 0 to 225 as shown in Matrix 1. Then, according to the normalisation method, the formula codes were applied.

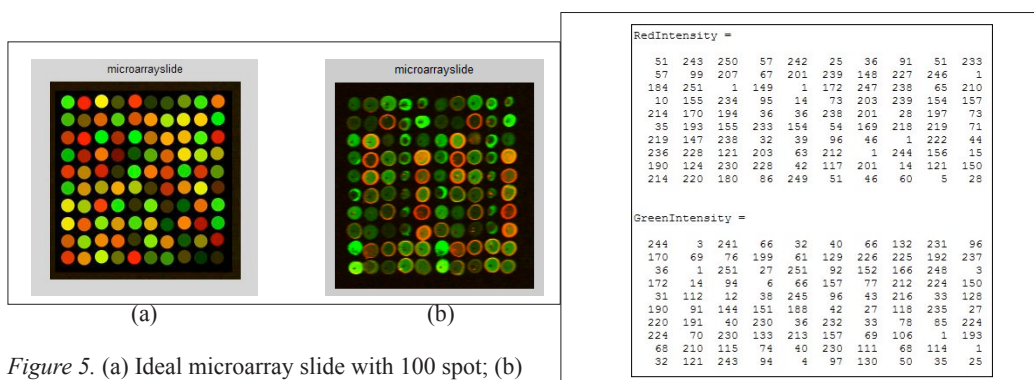


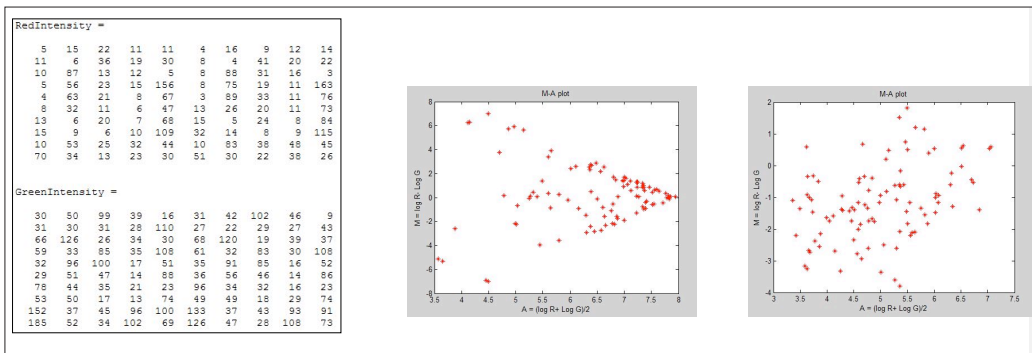
Figure 5. (a) Ideal microarray slide with 100 spot; (b) Real microarray slide with 100 spot;

Matrix 1. Original Intensity of the ideal spots

For Global Normalisation, loops were used to find the total of red and green intensities values for all 100 spots. Then taking the logarithm of the total value and subtracts it from the value of M according to Equation number 1 and 2. Similarly, in the Lowess method, mean of m values was calculated then subtracted, to be on the centre ($M=0$) according to equation number 3. However, Quantile normalisation is much different than the previous two methods, because it does not require calculation of A and M values. But it requires sorting the matrix in each column. Then taking the average in each row and finally put each new value in its original location as shown in Figure 5. Finally, Print-tip normalisation, A values (addition of logarithm) has been divided into four groups (<5 , <6 , <7 and else) and according to each group, mean value of M was taken and defined into variable call PT . After that, the PT value was subtracted from M according to its group. Next section will discuss the results of the various methods tested.

RESULT AND DISCUSSION

First of all, there is a different in the last result for all the four methods in terms of last intensities values and M - A displaying graphs. Global normalisation and Lowess share a similarity especially when we compare the difference between the green and the red intensity for the same spots. Similarly, Print-tip normalisation which has a similar graph but there is a different according to the interval groups. However, the results for quantile normalisation are fluctuating and the different is larger than all of the other normalisation methods. Normalisation results for the ideal and normal microarray slide are shown in matrix 1 and 2, and M - A graphs in Figure 6 and 7 respectively. As we saw in Matrix 1 above, there are red and green intensities for 100 spots as well as in Matrix 2 below. Thus, we have 4 matrixes with the size of (10×10) . The first and second for the red and green intensities of ideal image in Matrix 2 while the third and fourth for the red and green intensities for the slide image. Figure 6 and 7 depict M - A plots for ideal and slide image before any method of normalisation was performed. Thus, the illustrations will help us compare them with the next results of various normalisation methods.

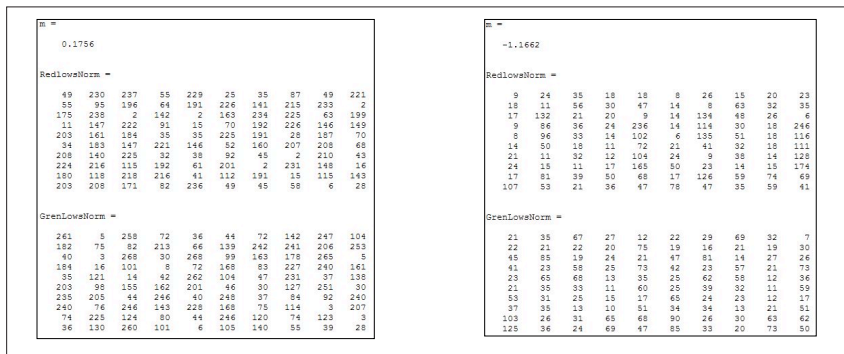


Matrix 2. Red and Green Intensity before norm of the slide image;

Figure 6. M-A plot before normalisation of the ideal image;

Figure 7. M-A plot before normalisation of the slide image

Matrixes 3 and Matrix 4 show the results of global normalisation for ideal and real DNA microarray slide. Firstly, they show k and c values, c is the logarithm of the total of red intensities over the total of green intensities (k) which is equal to 0.0274 in ideal image and -0.2358 , and that explains to us why the normalisation is important and how the variety of c increased for the real microarray slide image. Thus, the difference between the last and original results in the real microarray slide is larger.



Matrix 3. Red and Green Intensity for global norm of the ideal image

Matrix 4. Red and Green Intensity for global norm of real slide image

Lowess normalisation results for ideal and real DNA microarray slide are shown in Matrix 5 and 6. First it shows (m) values, m is the mean M value for 100 spots which equal to the difference between logarithms of red and green intensities for each spot separately. (m) is equal to 0.1756 in ideal image and -1.1662 explaining why the normalisation process is important and how does the variety of c increase for the real microarray slide image. Also, it is greater than c values (for global normalisation). The difference t between the last and original results in real microarray slide is larger than the different in global normalisation.

Image Normalisation Methods for DNA Microarray Data

<p>Matrix 5: Red and Green Intensity for global norm of the ideal image</p> <p>0.1756</p> <p>RedLowNorm =</p> <pre> 49 230 237 55 229 25 35 87 49 221 55 95 196 64 191 226 141 235 233 2 175 238 2 142 2 163 234 225 63 199 11 147 222 91 15 70 192 226 146 149 203 161 184 35 35 225 191 20 187 70 34 183 147 221 146 92 160 207 208 68 208 140 223 32 38 92 45 2 210 43 224 216 115 192 61 201 2 231 148 16 180 118 218 216 41 112 191 15 115 143 203 208 171 82 236 49 45 88 6 28 </pre> <p>GreenLowNorm =</p> <pre> 261 5 258 72 36 44 72 142 247 104 182 75 82 213 66 139 242 241 206 283 40 3 268 30 248 99 163 178 248 5 184 16 103 5 72 168 83 227 240 161 35 121 14 42 262 104 47 231 37 138 209 98 155 162 201 46 90 127 251 30 235 205 44 246 40 248 37 44 92 240 240 76 246 143 228 168 75 114 3 207 74 225 124 80 44 246 120 74 133 3 96 130 260 101 6 105 140 55 39 28 </pre>	<p>Matrix 6: Red and Green Intensity for global norm of real slide image</p> <p>-1.1662</p> <p>RedLowNorm =</p> <pre> 9 24 35 18 18 8 26 15 20 23 18 11 56 30 47 14 8 63 32 35 17 132 21 20 9 14 134 48 26 6 9 86 36 24 236 14 114 30 18 246 8 96 33 14 102 6 135 51 18 116 14 50 18 11 72 21 41 32 19 111 21 11 32 12 104 24 9 38 14 128 24 15 11 17 165 50 23 14 15 174 17 81 39 50 68 17 124 59 74 69 107 53 21 36 47 78 47 35 59 41 </pre> <p>GreenLowNorm =</p> <pre> 21 35 67 27 12 22 29 49 32 7 22 21 22 20 75 19 16 21 19 30 45 85 19 24 21 47 81 14 27 26 41 23 58 25 73 42 23 57 21 73 23 65 68 13 35 25 42 58 12 36 21 35 33 11 60 25 39 32 11 59 53 31 25 15 17 65 24 23 12 17 37 35 13 10 51 34 34 13 21 51 103 26 31 65 68 90 24 30 63 42 125 36 24 69 47 85 33 20 73 50 </pre>
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Matrix 5. Red and Green Intensity for global norm of the ideal image

Matrix 6. Red and Green Intensity for global norm of real slide image

Quantile normalisation results for ideal and real DNA microarray slide are shown in Matrix 7 and 8. Quantile normalisation method differs from global and Lowess normalisations in that it does not require fixed values of (c) or (m). Rather an average of the columns after sorting the matrix in each row as explained before in section 2. Thus, we can see in Matrix 8 that (67, 85, 124, 18 and so on) are repeated in each column of matrix QRN, and also the other values for QGN are similar in Matrix 8. There are 10 fixed numbers repeated in each column of each matrix.

<p>Matrix 7: R & G Intensity for Quant. Norm. of the ideal image</p> <p>QRN =</p> <pre> 67 229 243 67 229 18 50 124 50 243 85 18 149 85 211 243 124 192 243 18 124 243 18 192 18 192 243 211 67 229 18 85 211 149 50 85 229 229 124 211 192 124 124 50 67 229 192 67 192 149 50 149 67 243 192 67 149 149 211 124 229 67 229 18 85 124 67 18 229 85 243 211 50 211 149 211 18 243 149 50 149 50 192 229 124 149 211 50 85 192 211 192 85 124 243 50 85 85 18 67 </pre> <p>QGN =</p> <pre> 236 32 187 72 32 18 72 123 187 99 99 72 48 216 99 123 236 236 123 236 48 18 236 32 236 48 216 167 236 32 123 48 72 18 123 167 123 187 167 167 18 167 18 48 216 72 48 216 32 123 167 123 123 187 167 32 18 99 216 72 187 216 32 236 48 236 32 48 72 216 216 99 167 167 187 187 99 72 18 187 72 236 99 99 72 216 167 32 99 18 32 187 216 123 18 99 187 18 48 48 </pre>	<p>Matrix 8: R & G Intensity for Quant. norm of the real slide image</p> <p>QRN =</p> <pre> 9 16 38 23 9 9 16 9 28 9 38 6 78 42 14 14 6 78 42 14 23 78 14 28 6 16 55 38 38 6 14 42 42 38 78 23 38 14 14 78 6 55 28 14 38 6 78 42 16 38 16 23 9 6 28 38 23 16 23 28 42 9 23 9 42 42 9 28 6 42 55 14 6 16 55 55 14 6 9 55 28 38 55 78 23 28 42 55 78 23 78 28 16 55 16 78 28 23 55 16 </pre> <p>QGN =</p> <pre> 25 44 104 73 20 25 44 120 73 20 30 20 30 44 120 20 20 36 36 36 57 120 25 52 30 57 120 25 57 30 52 25 73 57 104 52 25 73 52 120 36 104 120 30 36 30 104 104 25 44 20 57 57 25 57 36 73 57 20 73 73 36 44 36 25 73 30 44 30 25 44 52 20 20 52 44 57 20 44 57 104 30 52 104 73 120 36 52 104 104 120 73 36 120 44 104 52 30 120 52 </pre>
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Matrix 7. R & G Intensity for Quant. Norm. of the ideal image

Matrix 8. R & G Intensity for Quant. norm of the real slide image

Finally, Print-tip normalisation gave the results for the red and green intensities for the ideal microarray image in Matrix 9 and real microarray slide in Matrix 10. M-A graphs for the results are displayed in Figure 8 and 9 respectively. PT values in Matrix 9 and 10 are represented by the normalisation values among the four intervals for each image. For example, in Matrix 9, PT equals -0.0664, 0.2457, 0.1445 and 0.2633. These values were subtracted from M (the different between logarithms of red and green intensities for each spot) according to the values of A for the same spot. These intervals are (<5, <6, <7 and else), so each interval has its own normalisation values; and that is why, at times, we can see the obvious different

between the normalised and un-normalised values in some intervals according to the values of PT. Besides that, Figure 9 represents the M-A plot for Print-tip normalization of real image slide which show more different from its original slide except by the values of PT especially in the first interval when $PT = -1.5263$ among the interval (A less than 5).

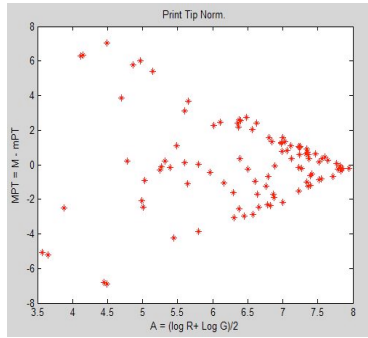


Figure 8. M-A Plot for Print tip norm of the Ideal Image

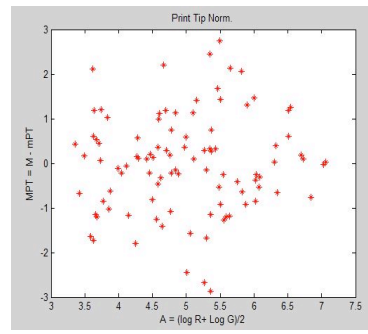


Figure 9. M-A Plot for Print tip norm of the Slide Image

PT =									
	-0.0664	0.2487	0.1445	0.2633					
RedPTNorm =									
50	250	230	54	232	24	34	88	50	214
56	96	198	65	193	220	137	209	226	3
176	258	3	143	3	165	227	219	61	216
11	144	215	89	16	71	195	220	142	145
208	187	178	94	36	219	193	28	189	71
35	178	143	214	142	51	162	200	201	67
201	136	228	32	37	89	44	3	204	43
217	218	112	187	61	195	3	224	161	15
182	115	211	210	40	108	185	14	117	155
205	202	166	83	230	50	45	56	7	30
GreenPTNorm =									
258	4	266	74	35	45	74	140	244	107
180	74	81	211	66	143	249	248	212	233
39	2	247	30	247	98	168	183	273	4
189	17	105	7	66	167	83	234	247	166
34	124	15	43	259	107	47	209	36	136
201	101	159	167	208	47	30	131	259	31
249	211	44	249	41	256	38	78	95	237
247	75	294	447	225	174	69	118	2	212
73	232	128	83	45	254	123	76	121	2
35	134	268	100	6	104	138	56	36	26

PT =									
	-1.5263	-0.9235	-0.6295	0.5579					
RedPTNorm =									
11	28	32	21	21	9	29	14	23	26
21	12	51	34	43	16	9	58	36	40
19	110	24	23	11	16	111	55	29	7
11	79	34	28	130	16	105	28	21	156
9	80	31	16	94	7	112	47	21	107
16	46	21	12	60	24	38	36	21	93
20	12	36	14	96	23	11	43	16	118
28	17	12	19	137	46	26	16	17	145
16	75	36	46	56	16	116	54	61	58
89	49	24	34	43	65	43	40	49	38
GreenPTNorm =									
19	31	73	24	11	19	26	75	28	6
19	19	24	18	81	17	14	22	17	26
40	103	16	21	19	41	98	12	24	23
36	25	63	22	133	37	24	61	19	133
20	78	74	11	38	22	74	68	11	39
18	38	29	9	72	22	42	28	9	70
58	27	22	15	18	71	21	20	11	18
32	31	11	9	61	37	30	12	15	61
112	28	34	71	82	98	28	32	76	74
150	39	21	75	51	103	35	18	88	54

Matrix 9. Red and Green Intensity for PT norm of the ideal image

Matrix 10. Red and Green Intensity for PT norm of real slide image

From the Matrixes and graphs discussed above, it can be observed the global and Lowes are almost similar; Print-tip, an advanced version of them, gave results that was close to Matrix 1 and 2. However, Quantile differed greatly than the correct one and its graphs fluctuate away from the goal. Furthermore, the graphs of real image Print-tip normalization shows the expected result for real slide image in Figure 9 due to the clustering around the straight line when ($M = 0$). These findings support the view of Smyth that the “print-tip loess normalization provides a well-tested general purpose normalization method which gives good results on a wide variety of arrays” and best combined with diagnostic plots of the data. When the diagnostic plots show that biases still remain in the data after normalization, additional steps such as quantile normalization of the arrays may be undertaken (Smyth et al., 2003).

CONCLUSION

In this paper, normalization is defined as a process to delete systematic error. Four most commonly used normalization algorithms such as Global, Lowess, Quantile and Print-tip were tested and compared to find the most suitable approach in a general normalization process. For that purpose, a Matlab code was built for each method for two slides; the ideal and real microarray slides. The results shown in the form of Matrix of red and green intensities and M-A graph show that Global, Lowess and Print-tip are more accurate in comparison with an ideal image result while Print-tip has the advantages than the other two especially in term of final graph shape.

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